

ERRATUM

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"This corrects the above-titled article. There was an error in the case-control label for a subset of samples. This was corrected and analyses were re-run. The thrust of the results and discussion did not change, but these results are more precise and corrected."

**NEXT-GENERATION ANALYSIS OF CATARACTS: DETERMINING
KNOWLEDGE DRIVEN GENE-GENE INTERACTIONS USING BIOFILTER, AND
GENE-ENVIRONMENT INTERACTIONS USING THE PHENX TOOLKIT***

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Investigating the association between biobank derived genomic data and the information of linked electronic health records (EHRs) is an emerging area of research for dissecting the architecture of complex human traits, where cases and controls for study are defined through the use of electronic phenotyping algorithms deployed in large EHR systems. For our study, cataract cases and controls were identified within the Marshfield Personalized Medicine Research Project (PMRP) biobank and linked EHR, which is a member of the NHGRI-funded electronic Medical Records and Genomics (eMERGE) Network. Our goal was to explore potential gene-gene and gene-environment interactions within these data for 527,953 and 527,936 single nucleotide polymorphisms (SNPs) for gene-gene and gene-environment analyses, respectively, with minor allele frequency > 1%, in order to explore higher level associations with cataract risk beyond investigations of single SNP-phenotype associations. To build our SNP-SNP interaction models we utilized a prior-knowledge driven filtering method called Biofilter to minimize the multiple testing burden of exploring the vast array of interaction models possible from our extensive number of SNPs. Using Biofilter, we developed 57,376 prior-knowledge directed SNP-SNP models to test for association with cataract status. We selected models that required 6 sources of external domain knowledge. We identified 13 statistically significant SNP-SNP models with an interaction with p-value < 1×10^{-4} , as well as an overall model with p-value < 0.01 associated with cataract status. We also conducted gene-environment interaction analyses for all GWAS SNPs and a set of environmental factors from the PhenX Toolkit: smoking, UV exposure, and alcohol use; these environmental factors have been previously associated with the formation of cataracts. We found a total of 782 gene-environment models that exhibit an interaction with a p-value < 1×10^{-4} associated with cataract status. Our results show these approaches enable advanced searches for epistasis and gene-environment interactions beyond GWAS, and that the EHR based approach provides an additional source of data for seeking these advanced explanatory models of the etiology of complex disease/outcome such as cataracts.

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1. Introduction

DNA biobanks coupled to electronic health records (EHR) have become a valuable resource for investigating the genetic architecture of complex traits, as EHR contain a wide array of medical information including billing codes and clinical laboratory measurements, often yielding a large sample size. Through carefully defining phenotypes, and using deployable algorithms that combine multiple sources of information in the EHR, cases and controls can be defined for association studies, such as defining age-related cataract cases and controls [1,2]. The Marshfield Personalized Medicine Research Project biobank (Marshfield PMRP) and linked EHR, used for the study described herein, is one such resource [3]. The Marshfield PMRP is a member of the NHGRI-funded electronic Medical Records and Genomics (eMERGE) Network, a network of similar biobanks coupled with EHR based data [4].

Cataracts are a leading cause of blindness globally [5], and are believed to arise from a combination of age, environmental factors, and heritable factors [6]. Thus, understanding the genetic etiology of cataracts, coupled with the effect of environment as a modifier, could have a profound impact on human health. For our study, algorithms proven for age-related cataract case identification [2] were deployed in the Marshfield PMRP EHR to identify 2580 cataract cases and 1367 controls, with further study details presented in Table 1. A total of 527,953 (gene-gene interactions) and 527,936 (gene-environment interactions) single nucleotide polymorphisms (SNPs) were available after PMRP genotyping coupled with quality control filtering and selection for SNPs with a minor allele frequency $> 1\%$.

Table 1. Marshfield Cataract Study Description

	Gene-Environment Analysis	Gene-Gene Analysis
Age	> 50	> 50
Ancestry	European American	European American
Total Samples	2,033	3,377
Cases	1,242	2,192
Controls	791	1,185
Males	821	1,408
Females	1,212	1,969
SNPs	527,936	527,953

Single SNP-phenotype associations are a dominant study design used in most genome-wide association studies (GWAS), however, more complex models that include interactions may more accurately describe the relationship between genetic variation and complex outcomes. Investigating all gene by gene (GxG), and in extension, all SNP by SNP (SNPxSNP) pairwise models is possible depending on the number of SNPs that have been genotyped. Unfortunately, the multiple hypothesis testing burden and risk of Type I error is inflated when investigating all pairwise models. A different approach can be used, utilizing prior biological knowledge methods directing model development. Thus, to investigate more complex models beyond single SNP-Phenotype associations for the Marshfield PMRP cataract dataset, we used the prior knowledge accessible through Biofilter 1.0 (a new implementation of Biofilter after the original description in [7]) to direct the investigation of pairwise GxG interaction models based on the following resources: the Kyoto Encyclopedia of Genes and Genomes (KEGG) [8], Reactome [9], Gene Ontology (GO) [10], the Protein families database (Pfam) [11], NetPath [12], Biological General Repository for Interaction Datasets (BioGrid) [13], and the Molecular INTeraction Database (MINT) [14]. Using the Biofilter, we developed 57,376 prior-knowledge directed SNPxSNP models to test for association with cataract status.

In addition, for this study we investigated gene-environment interactions (GxE), as there are clearly known environmental exposures that increase cataract risk, and when incorporated into analyses, may provide new models for the contribution of both environment and genetic architecture to cataracts. The Marshfield PMRP collected standardized Phenotypes and eXposures (PhenX) measures as a member of the PhenX Real-world, Implementation, SharingING (PhenX RISING) project. PhenX has the goal of defining standard phenotypic measures through a framework of measurement protocols via a web-based toolkit [15]. Environmental exposures such as smoking, sun exposure, and alcohol use, have been associated with increased cataract rates [16]. Thus we used 12 PhenX defined environmental exposures to investigate GxE interactions for the Marshfield PMRP cataract data focused on smoking, UV exposure, and alcohol use measures.

Through integrating EHR data, advanced bioinformatics tools, and PhenX, we can pursue advanced searches for epistasis and gene-environment interactions in genome-wide studies of common disease.

2. Methods

2.1. Marshfield EHR and Age-Related Cataract Case Identification

The Marshfield PMRP is a population based biobank with ~20,000 subjects, aged 18 years and older, enrolled in the Marshfield Clinic healthcare system in central Wisconsin [3]. DNA, plasma, and serum samples are collected at the time the enrollee completes a written informed consent document, with allowance for ongoing access to the linked medical records. PMRP participants also complete questionnaires, including responses regarding smoking history, occupation, and diet.

To identify cataract surgery cases aged 50 years and older within the PMRP, Current Procedural Terminology (CPT) codes in the Marshfield Clinic EHR were used. A research coordinator manually abstracted additional information to identify the eye affected, the type and severity of the cataract, and the level of visual acuity prior to the cataract surgery. This was also done to remove any cases with non-age related cataracts.

To identify individuals with diagnosed cataracts but without surgery, and to identify the type of cataract, International classification of diseases, 9th revision (ICD-9) codes and CPT codes were used, coupled with Natural Language Processing (NLP) and Intelligent Character Recognition (ICR) of free-text in the EHR. NLP and exclusion criteria were used to identify individuals with congenital and traumatic cataracts for omission from the study. Further details of the identification of cataract cases and controls and the efficacy of the EHR defined phenotyping can be found in Waudby et al., 2011 [2]. All total, the procedures used on the EHR identified 2,192 cases and 1,185 controls for gene-gene analysis and 1,242 cases and 791 controls for gene-environment analysis.

2.2. Genotyping

The eMERGE network and the Center for Inherited Disease Research (CIDR) at Johns Hopkins university performed the genotyping of the Marshfield PMRP samples, using the Illumina Human660W-Quadv1 A platform with total of 560,635 SNPs and 96,731 intensity-only probes. Bead Studio version 3.3.7 was used by CIDR for the genotyping calls. The total

cohort genotyped included 3947 samples from the Marshfield PMRP, 21 blind duplicates, and 85 HapMap controls. The HapMap concordance rate was 99.8% and the blind duplicate reproducibility rate was 99.99%. For quality control and data cleaning the eMERGE quality control (QC) pipeline developed by the eMERGE Genomics Working Group [17] was used. Any SNPs with a minor allele frequency > 1%, SNP call rate > 99%, Sample Call Rate > 99% were used in further analysis. After QC and allele frequency filtering using PLINK [18], a total of 527,953 and 527,936 SNPs were used for further gene-gene and gene-environment analyses, respectively.

2.3. PhenX

The standardized phenotypic and environmental consensus measures for Phenotypes and eXposures (PhenX) [15] were used to capture the environmental variables used in this study. The PhenX Toolkit (<https://www.phenxtoolkit.org/>) offers high-quality, well-established, standard measures of phenotypes and exposures for use in epidemiological studies.

The Marshfield PRMP is part of the PhenX RISING consortium, which is comprised of seven groups funded by the National Human Genome Research Institute (NHGRI) and the Office of Behavioral and Social Sciences Research (OBSSR) to incorporate PhenX (<https://www.phenxtoolkit.org/>) measures into existing population-based genomic studies.

For this initiative, Marshfield PRMP subjects with GWAS data who were alive with known, non-institutionalized addresses and who had given consent for re-contact were mailed a 32-page self-administered questionnaire that contained 35 PhenX measures across a range of phenotypic domains including alcohol and tobacco use questions (McCarty et al. 2012, *in preparation*). For this study, we considered 12 of these measures, shown in Table 2.

2.4. BioFilter 1.0

For the SNPxSNP analysis, Biofilter 1.0 was used. Biofilter has been upgraded from the initial Biofilter 0.5 [7], with the addition of more data sources, improved the handling of data, and the development of an eternal database for prior knowledge called the Library of Knowledge Integration (LOKI). Biofilter 1.0 and LOKI are freely available for non-commercial research institutions. For full details see: <http://ritchielab.psu.edu/ritchielab/software>.

Biofilter 1.0 utilizes prior biological knowledge through accessing the data of several publically available biological information databases, all compiled within the LOKI database developed specifically for Biofilter. The data sources selected for Biofilter contain information on networks, connections, and/or pathways that establish relationships between genes and gene products. Biofilter is a “gene based” approach, thus all the region information (such as genes) and position information (such as SNPs) are mapped to genes within LOKI.

The following sources that are compiled within LOKI were used for the Biofilter model building: the Kyoto Encyclopedia of Genes and Genomes (KEGG) [8], Reactome [9], Gene Ontology (GO) [10], the Protein families database (Pfam) [11], and NetPath [12], Biological General Repository for Interaction Datasets (BioGrid) [13], and the Molecular INTERaction Database (MINT) [14]. The database source used in LOKI solely for the purpose of mapping SNPs to genes is the National Center for Biotechnology (NCBI) dbSNP [19] database.

Table 2. The PhenX measures used for this study

PhenX Measure	Survey Question
PX030301 Alcohol 30Day Frequency	During the past 30 days, on how many days did you drink one or more drinks of an alcoholic beverage?
PX030301 Alcohol 30Day Quantity	During the past 30 days, how many drinks did you usually have each day?
PX030602 Cigarette Smoking 100	Have you smoked at least 100 cigarettes in your entire life?
PX030602 Cigarette Smoking Current	Do you now smoke cigarettes every day, some days, or not at all?
PX030602 Cigarette Smoking Everyday 6Month	Have you EVER smoked cigarettes EVERY DAY for at least 6 months?
PX030802 Everyday Smoker Quantity 1Day	On the average, about how many cigarettes do you now smoke each day?
PX030802 Someday Smoker Days 1Month	On how many of the past 30 days did you smoke cigarettes?
PX030802 Someday Smoker Quantity 1Day	On the average, on those days, how many cigarettes did you usually smoke each day?
PX030802 Former Smoker Smoking 6Month	Have you EVER smoked cigarettes EVERY DAY for at least 6 months?
PX030802 Former Smoker Quantity 1DayA	When you last smoked every day, on average how many cigarettes did you smoke each day?
PX030802 Former Smoker Quantity 1DayB	When you last smoked fairly regularly, on average how many cigarettes did you smoke each day?
PX061301 Weekend Sun Hours Last Decade	On a typical weekend day in the summer, about how many hours did you generally spend in the mid-day sun in the past ten years?

The following process was used within Biofilter 1.0 to develop the SNPxSNP models used in prior knowledge directed association testing. Figure 1 shows a simplified example of how the Biofilter 1.0 model generation process works. First, the input list of SNPs are mapped to genes within Biofilter. Next, comprehensive pairs of genes that are all terminal leaves of the graph for Pathway 1 in Source 1, and Pathway 2 in Source 1 are generated, only for genes that contain SNPs in the input list of SNPs.

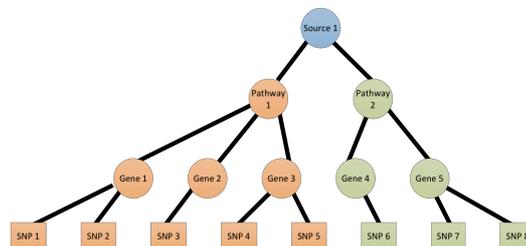


Figure 1. Simplified model for one Biofilter 1.0 database source with 2 pathways, 5 genes, and 8 SNPs

Implication scores are used in Biofilter to give each pairwise model a “score” indicating how many sources have that connected pair of genes represented, the higher the implication score, the more sources have indicated a connection between a pair of genes. The implication index is a measure of the number of data sources providing evidence of an interaction, a sum of the number of data sources supporting each of the two genes and the connection between them. In the case of

our simplified example, for Genes 1-5, that all contain SNPs within the input list, the following pairwise Gene-Gene models would result, each with an implication score of 1:

- Gene 1 – Gene 2
- Gene 1 – Gene 3
- Gene 2 – Gene 3
- Gene 4 – Gene 5

This process continues through all other sources used for Biofilter. Each time a pairwise combination of genes is found in another source (such as the pair Gene1-Gene2), the implication score for that pairwise model will be increased by 1. Lastly, the G-G models are broken into all pairwise combinations of SNPs across the genes, *within P1 or P2*. The SNP-SNP models would look like the following:

- SNP1-SNP3
- SNP1-SNP4
- SNP1-SNP5
- SNP2-SNP3
- SNP2-SNP4
- SNP2-SNP5
- SNP3-SNP5
- SNP3-SNP4
- SNP6-SNP7
- SNP6-SNP8

This same process was used within Biofilter 1.0 to develop the SNPxSNP models used for our prior knowledge directed association testing. First, the 527,953 SNPs were mapped to their corresponding genes. Next, the genes corresponding to the SNPs of the dataset were mapped to the gene-relationship graphs for each LOKI source used. After this mapping process, gene pairs were exhaustively generated for each occurrence of two genes within a single pathway and single source. Implication scores were calculated for the pairwise models. After the gene-gene models were developed in Biofilter, the models were divided into exhaustive SNP-SNP pairs for association testing.

Table 3 indicates the number of models that were found at each implication score cutoff. An implication index cutoff of 4 actually incorporates all possible pairwise models for all SNPs we had for this study, a total of 603,032 models. We found an implication score cutoff of 6 resulted in a balance between a large group of models for exploration (57,376 models), but still maintained a very computationally feasible set of associations to investigate, limiting our type 1 error rate more than using all exhaustive pairs of SNP-SNPs or some of the less stringent implication score cutoffs. With a requirement for an implication index of 6, as we had in this study, the gene-gene relationship or known interaction had to be found in nearly all of the data sources we used within LOKI.

Table 3. Number of Resulting Models for Each Implication Score Cutoff

Implication Index Cutoff	Number of Models
4	603,032
5	337,113
6	57,376
7	2479

2.5. Statistical Analysis

For the SNPxSNP models generated through the use of Biofilter, PLATO [20] was used to determine the significance of the interaction via likelihood ratio test (LRT) of the full versus

reduced models, using logistic regression, where the full model was: SNP1 + SNP2 + SNP1*SNP2 and the reduced model was: SNP1 + SNP2 for all of the pairwise sets of SNPs generated by Biofilter with an implication index of 6. For the GxE (SNPxE) models, the same methods were employed using PLATO; however the full model was: SNP1 + ENV1 + SNP1*ENV1 and the reduced model was: SNP1 + ENV1 for all the possible unique SNPxE pairs, from the set of 527,936 SNPs and the PhenX variables described earlier in methods. Again, the outcome was case control status for cataracts. The GGPlot2 [21] package in R was used for Figure 2.

3. Results

3.1. GxE Results

Figure 2 shows a Manhattan plot of the association results for the PhenX GxE models that had interaction with LRT p-values $\leq 1 \times 10^{-4}$, a total of 782 models exhibited an interaction with a p-value $\leq 1 \times 10^{-4}$ associated with cataract status. The top five GxE interaction results for each PhenX measure are also presented in Table 4, sorted by chromosome to highlight results similar across SNPs and regions for multiple PhenX measures. The measurement “Weekend Sun Hours Last Decade” a survey question asking “On a typical weekend day in the summer, about how many hours did you generally spend in the mid-day sun in the past ten years?” with the SNP rs6447541, located in an intron of *GABRI* on chromosome 4, with an association LRT p-value of 2.35×10^{-8} , was the most significant interaction found when compared to the other 12 PhenX measurements we used in our GxE analysis.

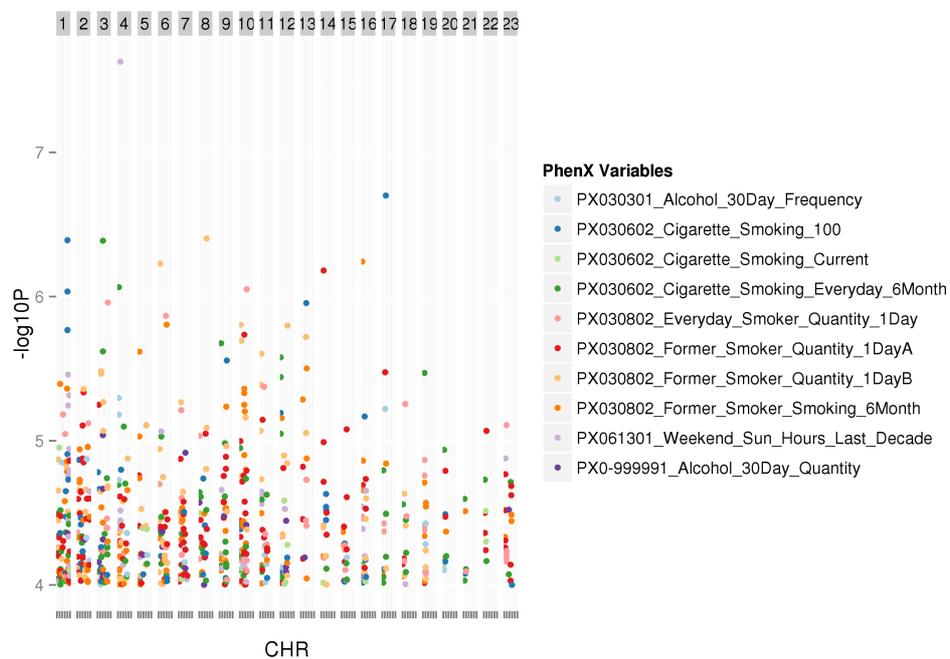


Figure 2. Manhattan plot of the association results for the GxE interaction models. Displayed are the results for the 12 PhenX measures that had interaction p-values $< 1 \times 10^{-4}$. Two PhenX variables did not have an interaction p-value less than 1×10^{-4} .

Table 4. Five most significant results for each PhenX measurement, sorted by chromosome and position

RSID	PhenX Variable	Chr:BP	P-value	Gene
rs7529518	PX030602_Cigarette_Smoking_100	1:200718422	1.71x10 ⁻⁶	<i>CAMSAP2</i>
rs2292097	PX030602_Cigarette_Smoking_100	1:200843768	9.23x10 ⁻⁷	<i>GPR25*</i>
rs10800745	PX030602_Cigarette_Smoking_100	1:200849676	4.06x10 ⁻⁷	<i>GPR25*</i>
rs11117581	PX061301_Weekend_Sun_Hours_Last_Decade	1:216613761	5.70x10 ⁻⁶	<i>USH2A*</i>
rs11117582	PX061301_Weekend_Sun_Hours_Last_Decade	1:216622208	3.48x10 ⁻⁶	<i>USH2A*</i>
rs10495409	PX061301_Weekend_Sun_Hours_Last_Decade	1:238255679	4.84x10 ⁻⁶	<i>MTND5P18</i>
rs607949	PX030602_Cigarette_Smoking_Current	1:43695708	1.12x10 ⁻⁵	<i>WDR65</i>
rs581503	PX030802_Former_Smoker_Smoking_6Month	1:61329593	4.03x10 ⁻⁶	<i>NFIA*</i>
rs11803470	PX030301_Alcohol_30Day_Frequency	1:95117783	1.42x10 ⁻⁵	<i>ABCD3*</i>
rs2587695	PX030802_Former_Smoker_Quantity_1DayA	2:120321817	4.62x10 ⁻⁶	<i>PCDP1</i>
rs262302	PX030301_Alcohol_30Day_Frequency	2:180931461	1.34x10 ⁻⁵	<i>CWC22*</i>
rs5994737	PX030602_Cigarette_Smoking_Current	22:33804804	3.07x10 ⁻⁵	<i>LARGE</i>
rs3846094	PX030602_Cigarette_Smoking_Everyday_6Month	3:101601159	2.40x10 ⁻⁶	<i>NFKBIZ*</i>
rs11720478	PX030602_Cigarette_Smoking_Everyday_6Month	3:101637806	4.10x10 ⁻⁷	<i>NFKBIZ*</i>
rs12495970	PX030802_Everyday_Smoker_Quantity_1Day	3:194928138	1.10x10 ⁻⁶	<i>XXYLTI</i>
rs11735349	PX030602_Cigarette_Smoking_Everyday_6Month	4:16506826	8.61x10 ⁻⁷	<i>LDB2</i>
rs157606	PX030301_Alcohol_30Day_Frequency	4:16699795	5.05x10 ⁻⁶	<i>LDB2</i>
rs283018	PX030301_Alcohol_30Day_Frequency	4:16740168	6.61x10 ⁻⁶	<i>LDB2</i>
rs6447541	PX061301_Weekend_Sun_Hours_Last_Decade	4:47215939	2.35x10 ⁻⁸	<i>GABRB1</i>
rs16888770	PX030802_Former_Smoker_Smoking_6Month	5:21586180	2.41x10 ⁻⁶	<i>GUSBP1</i>
rs13183503	PX030602_Cigarette_Smoking_Current	5:81515885	4.04x10 ⁻⁵	<i>ATG10</i>
rs9376419	PX030802_Everyday_Smoker_Quantity_1Day	6:139801295	1.36x10 ⁻⁶	<i>TXLNB</i>
rs3798756	PX030802_Former_Smoker_Smoking_6Month	6:152529260	1.56x10 ⁻⁶	<i>SYNE1</i>
rs3094549	PX030802_Former_Smoker_Quantity_1DayB	6:29355148	5.91x10 ⁻⁷	<i>OR12D2*</i>
rs4712006	PX061301_Weekend_Sun_Hours_Last_Decade	6:52245415	8.61x10 ⁻⁶	<i>PAQR8</i>
rs3889488	PX030802_Former_Smoker_Quantity_1DayB	8:141544748	3.96x10 ⁻⁷	<i>AGO2</i>
rs6987670	PX030602_Cigarette_Smoking_Current	8:9883177	3.18x10 ⁻⁵	<i>MSRA*</i>
rs10968388	PX030602_Cigarette_Smoking_Everyday_6Month	9:28210699	2.11x10 ⁻⁶	<i>LINGO2</i>
rs9783135	PX030802_Everyday_Smoker_Quantity_1Day	10:129937722	8.90x10 ⁻⁷	<i>MKI67*</i>
rs12360020	PX030802_Former_Smoker_Quantity_1DayB	10:15264322	1.57x10 ⁻⁶	<i>FAM171A1</i>
rs2820100	PX030802_Former_Smoker_Quantity_1DayA	10:84491173	1.84x10 ⁻⁶	<i>NRG3</i>
rs6592528	PX030802_Everyday_Smoker_Quantity_1Day	11:73377350	4.22x10 ⁻⁶	<i>PLEKHB1*</i>
rs4944859	PX030802_Everyday_Smoker_Quantity_1Day	11:73424135	4.22x10 ⁻⁶	<i>RAB6A</i>
rs7977795	PX030802_Former_Smoker_Quantity_1DayB	12:132096632	1.59x10 ⁻⁶	<i>SFSWAP*</i>
rs7972947	PX030602_Cigarette_Smoking_Everyday_6Month	12:2170433	2.64x10 ⁻⁶	<i>CACNA1C</i>
rs775474	PX030602_Cigarette_Smoking_Current	12:70075933	2.60x10 ⁻⁵	<i>BEST3</i>
rs680711	PX030602_Cigarette_Smoking_100	13:101814804	1.11x10 ⁻⁶	<i>NALCN</i>
rs4772995	PX030802_Former_Smoker_Smoking_6Month	13:109410933	3.15x10 ⁻⁶	<i>MYO16</i>
rs7983958	PX030802_Former_Smoker_Quantity_1DayB	13:96473682	1.90x10 ⁻⁶	<i>UGGT2</i>
rs1957480	PX030802_Former_Smoker_Quantity_1DayA	14:44397890	6.59x10 ⁻⁷	<i>X10IF4BP1*</i>

rs11644531	PX030802_Former_Smoker_Smoking_6Month	16:6008824	5.72x10 ⁻⁷	<i>RBFOX1</i> *
rs8075882	PX030301_Alcohol_30Day_Frequency	17:55469362	6.01x10 ⁻⁶	<i>MSI2</i>
rs1443269	PX030802_Former_Smoker_Quantity_1DayA	17:55894564	3.35x10 ⁻⁶	<i>MRPS23</i> *
rs9911607	PX030802_Former_Smoker_Quantity_1DayA	17:55895539	3.35x10 ⁻⁶	<i>MRPS23</i> *
rs7210514	PX030602_Cigarette_Smoking_100	17:67793814	1.99x10 ⁻⁷	<i>KCNJ16</i> *

Table Abbreviations: Chr = Chromosome; BP = Base pair location of SNP; RSID = SNP ID; P-value = P-value of the interaction; Gene = Gene symbol of gene is within or nearest to (*indicates nearest gene is listed)

3.2. GxG Results

The top Biofilter 1.0 derived GxG models are presented in Table 5. A total of 13 models had an LRT p-value $< 1 \times 10^{-4}$ and full model p-value < 0.01 . A total of 9 genes were in the thirteen models. Of these models, the most significant was for a model with *SOS1*, which encodes a guanine nucleotide exchange factor for RAS proteins, and *FYN*, which is a member of the protein-tyrosine kinase oncogene family.

Table 5. The 13 SNPxSNP models with an interaction p-value $< 1 \times 10^{-4}$ after association testing of the Biofilter derived pairwise models.

SNP1	Gene 1	SNP2	Gene 2	Interaction P-value
rs2888586	<i>SOS1</i>	rs706885	<i>FYN</i>	1.29x10 ⁻⁶
rs2888586	<i>SOS1</i>	rs17072912	<i>FYN</i>	2.14x10 ⁻⁶
rs2888586	<i>SOS1</i>	rs11964650	<i>FYN</i>	2.97x10 ⁻⁶
rs2888586	<i>SOS1</i>	rs9372313	<i>FYN</i>	6.32x10 ⁻⁶
rs17446875	<i>CDH2</i>	rs6121791	<i>CDH4</i>	2.64x10 ⁻⁵
rs9384805	<i>FYN</i>	rs11017910	<i>DOCK1</i>	2.67x10 ⁻⁵
rs11083252	<i>CDH2</i>	rs6121791	<i>CDH4</i>	4.39x10 ⁻⁵
rs13135792	<i>KIT</i>	rs10515074	<i>PIK3R1</i>	4.74x10 ⁻⁵
rs631428	<i>COL4A1</i>	rs3803231	<i>COL4A2</i>	6.67x10 ⁻⁵
rs613116	<i>COL4A1</i>	rs3803231	<i>COL4A2</i>	6.99x10 ⁻⁵
rs17704348	<i>FYN</i>	rs4751282	<i>DOCK1</i>	8.85x10 ⁻⁵
rs17446875	<i>CDH2</i>	rs1110359	<i>CDH4</i>	8.85x10 ⁻⁵
rs809193	<i>FYN</i>	rs11594969	<i>DOCK1</i>	9.64x10 ⁻⁵

4. Discussion

The results presented herein are an exploration of the use of multiple novel approaches for investigating gene and phenotype associations within EHR based data. We performed an analysis with PhenX derived measures, seeking GxE interaction models for the Marshfield Cataract data set. The majority of the significant interactions were found for smoking related measures. We did find some highly correlated PhenX measures with significant interactions for SNPs within similar regions, such as the results on chromosome 1 for SNPs rs2292097 and rs7529518, for smoking related phenotypes. Through searches in the NCBI catalog [22], as well as the National Center for Biotechnology (NCBI) dbSNP [19], these two SNPs, as well the SNP in our most significant GxE model, did not show previous GWA level significant associations for any phenotypes.

We also performed an exploratory analysis with Biofilter 1.0, an updated and improved implementation of the originally published Biofilter. The results are intriguing, and provide the basis for hypotheses that can be investigated further, highlighting how Biofilter results have a biological context that provide additional information for resulting models. Interestingly, three of the models that passed our significance cutoff contained two of the same genes, *FYN*, a member of

the protein-tyrosine kinase oncogene family implicated in cell growth, and *DOCK1*, dedicator of cytokinesis 1. These models as a whole implicate genes related to cell growth, the cell cycle, and epidermal growth.

We are currently developing Biofilter 2.0 which will include additional database sources and allow for the use of other position and region based information beyond SNPs and genes, such as copy number variation (CNV) data, evolutionary conserved regions, and regulatory regions, allowing users to incorporate additional sources of prior knowledge as well as utilize other sources of genetic variation measurement data, with a more user-friendly interface.

Our results provide more complex models for an association between genetic variation and cataract outcome, moving beyond the more standard SNP-phenotype associations. The models found we intend to investigate further and warrant additional investigation of the environment and genetic variables contributing to these more complex models. These bioinformatics approaches can be used with other datasets, to expand the investigation of the relationship between genetic architecture and phenotypic outcome. With these approaches that consider the complexity of the data and harness the power of novel bioinformatics tools, we will elucidate the missing heritability of complex traits.

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